

FACULTY OF ENGINEERING

DEPARTMENT OF GINNING AND TEXTILE ENGINEERING

PROGRAMME: BACHELOR OF SCIENCE IN TEXTILE ENGINEERING

RESEARCH PROJECT TITLE:

ANTIMICROBIAL TREATMENT OF COTTON FABRIC WITH BIDENS PILOSA LEAF EXTRACTS

Regno. BU/UG/2014/101

NAME: OSHABA JONAN

CONTACTS: 0777681685/0756117907

EMAIL:oshabajonan093@gmail.com

SUPERVISOR NAME: Madam, NAMUGA CATHERINE

A Project Report Submitted to The Department of Ginning and Textile Engineering in Partial Fulfilment of the Requirements for The Award of a Bachelor's Degree in Textile Engineering.

May 2018

ACKNOWLEDGEMENT

· · ·

I acknowledge the great power of God who made this research successful, and his protection throughout. I thank Almighty God for providing a way for me even in the most hopeless times especially in financial issues and providing guides.

The financing from my family especially my aunt Mrs. MUGISHA ANNE and her husband Mr. MUGISHA STEVEN made my journey easier in this research. Their love and encouragement has helped me to keep focused and hopeful for the best in the research work.

I also acknowledge the guidance of my supervisor Madam NAMUGA CATHERINE; whose unlimited guidance and help led to this research. I thank her for her role in organization of the work, tips and knowledge availed to me up to the end of research.

I acknowledge the opportunity given to me by UIRI (Uganda industrial research institute) management for allowing me to carry out my work. I also thank the staff for guiding me throughout the process. Great thanks go to Southern Range Nyanza Limited for providing me with the cotton samples that were used.

I acknowledge the Busitema university administration and staff for providing a suitable environment for research to go on smoothly. Great thanks go to the head of department Textile and Ginning, for his friendly heart, words of wisdom and guidance.

Lacknowledge the role of my friends in encouragements and giving me hope. Their hospitality and accommodation services enabled completion of the research.

DECLARATION

I **Oshaba Jonan** declare that the information contained in this report was written by me and has not been copied from anywhere else. It is the information I researched and results from the experiments or tests that I carried out. I declare that it has not been presented anywhere in any institution for the award of any academic certificate, diploma or degree.

NAME: OSHABA JONAN

REGNO: BU/UG/2014/101

BUSITEMA UNIVERSITY LIBRARY
CLASS No.:
ACCESS NO. FET OLL 80

APPROVAL I present this report to the following persons for approval.

SUPERVISOR

NAME: Madam NAMUGA CATHERINE
SIGNATURE:
DATE:

HEAD OF DEPARTMENT

NAME; Dr. NIBIKORA ILDPHONSE

SIGNATURE:

DATE:

DEDICATION

 F_{ij}^{*}

453

This research is dedicates to my aunt Mrs. MUGISHA ANNE and her husband Mr. MUGISHA STEVEN; all of whom have contributed tremendously towards the completion of this research, both spiritually and financially.

I also dedicate this report to my brother and sisters, all my aunts and family friends; who have prayed for me and wished me all the best.

I dedicate this report to my friends; KATO PETER, KAMARA, NIKUZE DEREK and all my classmates. Their company, advice and encouragement have been of much help.

I dedicate this research to all my lecturers at Busitema university especially the head of department Textile and ginning; Dr. NIBIKORA ILDPHONSE, who have played part in my learning. They have been open in delivering knowledge.

I dedicate this research to my supervisor Madam NAMUGA CATHERINE; who has been the backbone of all the processes in this research.

I dedicate the research to; the chemistry staff at UIRI especially Mr. WILBER and Ms. JUSTINE, the microbiology staff UIRI Madam CATHERINE and Madam JIN, including the entire UIRI administration.

I dedicate this research to Madam MONIC, Mr. FRANCIS from the chemistry lab Southern Range Nyanza Ltd and the administration.

ABSTRACT

The study covers leaves of bidens pilosa with the bioactive compounds extracted using solvents of ethanol, water and acetone which were later applied to cloth for antimicrobial activity test. Soxhlet extraction method was used. The number of phytochemicals whose test was positive were higher for organic extracts than for aqueous extracts. Flavonoids, glycosides, tannins, phenols, terpenoids tested positive for ethanol and acetone extracts while flavonoids and glycosides tested positive in water extract. The yield of the extract was 16% for water and 5.7%; for ethanol, Only extracts of water and ethanol were applied on cloth for study of the antimicrobial property. The agar diffusion method gave 17mm diameter as zone of inhibition for water extract against staphylococcus aureus with organic extracts exhibiting no antimicrobial activity while ciprofloxacin used as the standard drug had 39mm inhibition zone. The minimum inhibitory concentration test against S. aureus carried out using the broth dilution method showed 20mg/ml for ethanol and acetone and 10mg/ml for water extract. The extracts were applied to the bleached cotton fabric by dipping in liquor ratio 20 and drying at 70°c. Curing was done at 150°c. Antimicrobial tests were carried out for treated cotton cloth against staphylococcus aureus (S. aureus) and Escherichia coli (E. coli) using the disc diffusion method for cloth. The cloth exhibited higher antimicrobial activity against S. aureus and very little activity against E. coli. The antimicrobial activity; against S. aureus, of water extract was high when the cloth of more concentration (27%) was used. The zone of inhibition was 9.5mm. The ethanol extract on cloth showed negligible effect. Extracts from bidens pilosa leaves when incorporated in cotton cloth show promising potential against S. aureus, specifically aqueous extracts.

LIST OF FIGURES

.

Figure 1 shows Shoot parts of bidens pilosa plant
Figure 2 shows drying process; left if the undried leaves and right is the dried leaves
Figure 3 Blender and the crushed leaves respectively
Figure 4 Powdered leaves after sieving
Figure 5 Left is soxhlet apparatus, next is gel for of the extract after evaporating the solvent
Figure 6 shows series of agar plates containing wells of extracts
Figure 7 Broth dilution tubes, agar nutrient plates respectively
Figure 8 the photo from left shows; cloth treated with extracts of ethanol 5.5%, water 5.5%, water 27%,
and untreated cloth respectively
Figure 9 shows E. coli and S. aureus plates for agar diffusion method. (E-ethanol, W-water, S-standard,
A-acetone)
Figure 10 agar plates showing the bacterial growth after incubation for 24h. From left, the columns are of acetone, ethanol and water respectively. The concentration is in decreasing order up along the column of
plates
Figure 11; shows the zone of inhibition for sample discs W1 and W2 at dilutions of 10°, 10 ⁻¹ for S. aureus
bacteria
Figure 12, E. coli, S. aureus agar plates respectively with fabric discs on top of nutrient agar
Figure 13 E. coli Figure 14 test for glycosides
Figure 14 positive test for flavonoids in water extract Figure 15 S. aureus bacteria

LIST OF ABBREVIATIONS AND ACCRONYMS

UIRI-Uganda Industrial Research Institute

E. coli- Escherichia coli

S. aureus- Staphylococcus aureus

m-Meter

ml- mil-liter

MIC- minimum inhibitory concentration

TTO- Tea tree oil

SFE –Supercritical fluid extraction

HCl -hydrochloric acid

FeCl₃- iron (III) chloride

H₂SO₄-Sulphuric acid

х.

NaOH- Sodium hydroxide

Contents ACKNOWLEDGEMENT
DECLARATION
APPROVAL
DEDICATIONv
ABSTRACT
LIST OF FIGURES
LIST OF ABBREVIATIONS AND ACCRONYMS
1.0. CHAPTER ONE
1.1. INTRODUCTION
1.2. BACKGROUND/LITERATURE REVIEW
1.2.1. Antimicrobial textiles
1.2.2. Mechanism of bioactive agents contained in antimicrobial textiles
1.2.3. Antimicrobial agents applied in textiles
1.2.4. Plants as antimicrobial active agent-sources
1.2.5. Extraction methods used for plant based compounds
1.2.6. Solvents used in extraction of plant constituent compounds
1.2.7. Phytochemistry of bidens pilosa
1.2.8. Separation of phytochemicals
1.2.9. Methods used for incorporating antimicrobial agents in the textile
1.2.10. Test of Antimicrobial Activity on textiles
1.3. PROBLEM STATEMENT
I.4. OBJECTIVES
1.4.1. Main objective;
1.4.2. Specific objectives:
1.5. Scope
1.6. Justification
2. CHAPTER TWO
2.1. Materials and methods/methodology;
2.1.1. Choice of plant;

2	2.1.2. Drying
2	2.1.3. Crushing and grinding
2	2.1.4. Sieving
2	2.1.5. Extraction of bioactive compound of bidens pilosa,
2	2.1.6. Phytochemical screening, qualitative analysis
2	2.1.7. Agar well diffusion method21
2	2.1.8. Determination of minimum inhibitory concentration (MIC)
2	2.1.9. Application of extracts on cloth
2	2.1.10. Antimicrobial Tests for cotton textile
3.0 CI	HAPTER 3
3.1	. RESULTS AND DISCUSSION
3	3.1.1. Yield of plant extract
3	3.1.2. Results for phytochemical screening, qualitative analysis
1	3.1.3. Results from agar diffusion test (Zone of inhibition)
3	3.1.4. Results for Minimum inhibitory concentration (MIC) for S. aureus
	3.1.5. Results for antimicrobial activity (semi-quantitative test), DIN EN ISO 20645 (agar plate diffusion test)
4.0. C	CONCLUSION
5.0. R	ECOMMENDATIONS
Refer	ences
APPE	NDICIES
Sch	hedule and budget

.

1.0. CHAPTER ONE

1.1. INTRODUCTION

There is increased need for functional textiles which perform different tasks other than only providing cover to humans, or in other applications. A lot of functional finishes have been developed and one of the tasks for these finishes in textiles is the antimicrobial activity and antioxidant activity.

Antimicrobial finishing is a chemical treatment for textiles that provides the ability to destroy or inhibit the growth of microscopic organisms. Antimicrobial textiles are rapidly advancing for use in various fields like industries, medical, engineering, agriculture and food. Textile fibres with built-in antimicrobial properties will also serve the purpose alone or in blends with other fibres. They are referred to as bioactive fibres. Bioactive fibres find application enormous areas such as sanitary materials, dressing materials, surgical threads, materials for filtration of gases and liquids, air conditioning and ventilation, constructional materials, special materials for food industry, pharmaceutical industry, footwear industry, clothing industry, automotive industry(Sun, 2016a). Antimicrobial textiles are rapidly advancing for use in various fields like industries, medical, engineering, agriculture and food(Shahid and Mohammad, 2013). Textile fibres with built-in antimicrobial properties will also serve the purpose alone or in blends with other fibres. They are referred to as bioactive fibres. Later, the home textiles, such as, curtains coverings, and bath mats came with antimicrobial finish. The application of the finish is now extended to textiles used for outdoor, healthcare sector, sports and leisure. Antimicrobial finishing technologies include pad drying, crosslinking, sol-gel technology, plasma technology, nanotechnology. Microencapsulation and application of cyclodextrins give slow release of the finishes and ensure durability in the textile. (Joshi, Ali and Purwar, 2009, Shahid and Mohammad, 2013)

Due to the problems associated with use of synthetic chemicals, especially pollution to environment and toxicity to humans, there is need for alternative biodegradable finishing agents *(Sathianarayanan et al., 2010)*. Natural biopolymers are the only suitable and renewable products. The antimicrobial activity of few plants species out of about 250,000-500,000 known plant species, has been under study (Sun, 2016b). This is attributed to their use as traditional medicines. One of the most used medicinal plants is bidens pilosa, which is a common species with in the tropics. It grows to an average height of about 0.6m and maximum height of 1.5m and is an annual plant. It origin is known to be South America. *(Science, 2016, Cortés-rojas, Chagas-paula and Fernando, 2013)*

The project covers 'cotton textile treated with antibacterial plant extracts'. The project shows bidens pilosa as the bioactive agent source. The whole process rests on the idea of functional finishes for antimicrobial textiles.

References

Afolayan, A. J. (2009) 'Screening the root extracts from Biden pilosa L. var. radiata (Asteraceae) for antimicrobial potentials', (September).

Al-deyab, S. S. (2015) 'Highly-effective-antibacterial-textiles-', *Carbohydrate Polymers*. Elsevier Ltd., 86(2), pp. 936–940. doi: 10.1016/j.carbpol.2011.05.048.

Arunkumar, S. and Muthuselvam, M. (2009) 'Analysis of phytochemical constituents and antimicrobial activities of Aloe vera L. against clinical pathogens', *World Journal of Agricultural Sciences*, 5(5), pp. 572–576. Available at:

http://www.idosi.org/wjas/wjas5(5)/9.pdf.

Ayoola, G. A. et al. (2008) 'Phytochemical Screening and Antioxidant Activities of Some Selected Medicinal Plants Used for Malaria Therapy in Southwestern Nigeria'.

Barimah, J. et al. (2017) 'Effect of Drying Methods on Phytochemicals, Antioxidant Activity and Total Phenolic Content of Dandelion Leaves', 5(4), pp. 136-141. doi: 10.12691/ajfn-5-4-4.

Bartolome, A. P., Villaseñor, I. M. and Yang, W. (2013) 'Bidens pilosa L. (Asteraceae): Botanical Properties, Traditional Uses, Phytochemistry, and Pharmacology', 2013.

Chemistry, C. (2016) 'Cold plasma treatment of cotton and viscose fabrics impregnated with essential oils of lavandula angustifolia and melaleuca alternifolia', 50, pp. 711–719.

Chien, S. et al. (2009) 'Phytochemistry Anti-diabetic properties of three common Bidens pilosa variants in Taiwan', *Phytochemistry*. Elsevier Ltd, 70(10), pp. 1246–1254. doi: 10.1016/j.phytochem.2009.07.011.

Cortés-rojas, D. F., Chagas-paula, D. A. and Fernando, B. (2013) 'Bioactive compounds in Bidens pilosa L. populations : a key step in the standardization of phytopharmaceutical preparations', *Revista Brasileira de Farmacognosia - BrazilianJournal of Pharmacognosy*. Elsevier, 23(1), pp. 28–35. doi: 10.1590/S0102-695X2012005000100.

Cushnie, T. P. T. and Lamb, A. J. (2005) 'Antimicrobial activity of flavonoids', 26, pp. 343-356. doi: 10.1016/j.ijantimicag.2005.09.002.

Deba, F. et al. (2008) 'Chemical composition and antioxidant, antibacterial and antifungal activities of the essential oils from Bidens pilosa Linn. var. Radiata', Food Control, 19(4), pp. 346–352. doi: 10.1016/j.foodcont.2007.04.011.

El-kamali, H. H. and Elshikh, A. A. (2015) 'Preliminary Phytochemical Screening of 27 Plants Species Use in Ethnoveterinary in Khartoum State, Sudan', 5(2), pp. 48–52. doi: 10.5923/j.als.20150502.03.

Gupta, V. (2016) 'UV Protective and Antibacterial Finish on Cotton Using Plant Extracts D o c t o r o f P h i l o s o p h y i n T e x t i l e a n d A p p a r e l D e s i g n i n g'.

Hassan, K. A. et al. (2011) 'Wound healing potential of the ethanolic extracts of Bidens pilosa and Ocimum suave', 5(February), pp. 132–136. doi: 10.5897/AJPP10.266.

Hooda, S. et al. (2013) 'EFFECT OF LAUNDERING ON HERBAL FINISH OF COTTON', 3(4), pp. 35-42.

Jayapriya, S. *et al.* (2013) 'TEXTILE ANTIMICROBIAL TESTING AND STANDARDS', 4(1), pp. 1–10.

Joshi, M., Ali, S. W. and Purwar, R. (2009) 'Ecofriendly antimicrobial finishing of textiles using bioactive agents based on natural products', 34(September), pp. 295-304.

Krishnaiah D et al. (2009) 'Studies on phytochemical constituents of six Malaysian medicinal plants', Journal of Medicinal Plants Research, 3(2), pp. 67-72. Available at:

http://www.academicjournals.org/JMPR.

Kviecinski, M. R. *et al.* (2011) 'The Journal of Supercritical Fluids SFE from Bidens pilosa Linné to obtain extracts rich in cytotoxic polyacetylenes with antitumor activity', *The Journal of Supercritical Fluids.* Elsevier B.V., 56(3), pp. 243–248. doi: 10.1016/j.supflu.2010.12.011.

Lee, C. et al. (2013) 'Correlations of the components of tea tree oil with its antibacterial effects and skin irritation', *Journal of Food and Drug Analysis*. Elsevier Ltd, 21(2), pp. 169–176. doi: 10.1016/j.jfda.2013.05.007.

Mandal, V., Mohan, Y. and Hemalatha, S. (2007) 'PHCOG REV .: Review Article Microwave Assisted Extraction – An Innovative and Promising Extraction Tool for Medicinal Plant Research', 1(1).

Maobe, M. A. G. et al. (2013) 'Preliminary Phytochemical Screening of Eight Selected Medicinal Herbs Used for the Treatment of Diabetes, Malaria and Pneumonia in Kisii Region, Southwest Kenya', 5(1), pp. 1–6. doi: 10.5829/idosi.ejas.2013.5.1.65127.

Mudau, F. N. and Ngezimana, W. (2014) 'Effect of Different Drying Methods on Chemical Composition and Antimicrobial Activity of Bush Tea (Athrixia phylicoides)', pp. 1011–1014.

Nn, A. (2015) 'Medicinal & Aromatic Plants A Review on the Extraction Methods Use in Medicinal Plants, Principle, Strength and Limitation', 4(3), pp. 3–8. doi: 10.4172/2167-0412.1000196.

Paulo, S. (2014) 'In vitro SCREENING ANTIBACTERIAL ACTIVITY OF Bidens pilosa LINNÉ AND Annona crassifiora MART . AGAINST OXACILLIN RESISTANT Staphylococcus aureus (ORSA) FROM THE AERIAL', 56(4), pp. 333–340. doi: 10.1590/S0036-46652014000400011.

Prakash, O., Kumar, A. and Kumar, P. (2013) 'Anticancer Potential of Plants and Natural Products : A Review', 1(6), pp. 104–115. doi: 10.12691/ajps-1-6-1.

*Pressurized hot water extraction (PHWE) and chemometric fingerprinting of phytochemicals from ... (2017), (September 2016). doi: 10.13140/RG.2.2.25633.81765.

Rojas, J. J. *et al.* (2006) 'BMC Complementary and Screening for antimicrobial activity of ten medicinal plants used in Colombian folkloric medicine : A possible alternative in the treatment of non-nosocomial infections', 6, pp. 1–6. doi: 10.1186/1472-6882-6-2.

Sathianarayanan, M. P. et al. (2010) 'Antibacterial finish for cotton fabric from herbal products', 35(March), pp. 50-58.

Science, F. (2016) 'Plant collection and extraction of essential oils'. doi: 10.1002/fsn3.330. Shahid, M. and Mohammad, F. (2013) 'Green Chemistry Approaches to Develop Antimicrobial Textiles Based on Sustainable Biopolymers \Box A Review'.

Silva, M. S., Athayde-filho, P. F. De and Barbosa-filho, J. M. (2011) 'Compilation of Secondary Metabolites from Bidens pilosa L.', pp. 1070–1102. doi: 10.3390/molecules16021070.

Sun, G. (ed.) (2016a) Antimicrobial Textiles. Cambridge: Woodhead Publishing Limited. Sun, G. (ed.) (2016b) Antimicrobial Textiles. Cambridge: Woodhead Publishing Limited. Available at: http://store.elsevier.com/.

Sundararajan, P. et al. (2003) 'Studies of anticancer and antipyretic activity of Bidens pilosa whole plant', (October), pp. 27-30.

Taylor, P. et al. (2012) 'The Journal of The Textile Institute Antibacterial effects of two Tunisian Eucalyptus leaf extracts on wool and cotton fabrics', (July 2014), pp. 37-41. doi: 10.1080/00405000.2012.670946.

Tradit, A. J. et al. (2011) 'EXTRACTION, ISOLATION AND CHARACTERIZATION OF BIOACTIVE COMPOUNDS FROM PLANTS ' EXTRACTS Institute for Research in Molecular Medicine (INFORM), Universiti Sains Malaysia, Minden 11800', 8, pp. 1–10. Ubillas, R. P. et al. (2000) 'Antihyperglycemic Acetylenic Glucosides from Bidens pilosa', 66, pp. 82–83.

.