Prediction of Ebolavirus Genomes Encoded MicroRNA-Like Small RNAs Using Bioinformatics Approaches

Yue Teng, Zhe Xu, Jin Yuan, Xiaoping An, Jiangman Song and Dan Feng

Additional information is available at the end of the chapter

http://dx.doi.org/10.5772/62944

Abstract

Recent findings revealed that certain viruses encoded microRNA-like small RNAs using the RNA interference machinery in the host cells. However, the function of these virusencoded microRNA-like small RNAs remained unclear, and whether these microRNAlike small RNAs were involved in the replication of the virus and viral infection was still disputable. In this chapter, the negative-sense RNA genome of Ebola virus (EBOV) was scanned using bioinformatics tools to predict the EBOV-encoded microRNA-like small RNAs. Then, the potential influence of viral microRNA-like small RNAs on the viral immune evasion, host cellular signaling pathway, and epigenetic regulation of antiviral defense mechanism were also detected by the reconstructed regulatory network of target genes associated with viral encoded microRNA-like small RNAs. In this analysis, EBOV-encoded microRNA-like small RNAs were proposed to inhibit the host immune response factors, probably facilitating the evasion of EBOV from the host defense mechanisms. In conclusion, systematic investigation of microRNA-like small RNAs in EBOV genome may shed light on the underlying molecular mechanisms of the pathological process of Ebola virus disease (EVD).

Keywords: Ebolavirus, virus-encoded miRNAs, microRNAs, bioinformatics, NF-kB, TNF

1. Introduction

Zaire Ebola virus (ZEBOV) has the highest case-fatality rate with an average of approximately 83% over the past 27 years [1]. Its first outbreak took place on August 26, 1976, in Yambuku [2], and the virus was also responsible for the 2014 West Africa outbreak, which was the largest EBOV outbreak in record [3–6]. Moreover, neither antiviral drugs nor effective treatment was available for EBOV or Ebola virus disease (EVD) at that time [7, 8]. MicroRNAs originate from a wide variety of primary transcripts (pri-miRNAs) that are generated by RNA polymerase II (pol II) in all eukaryotes [9] or by RNA polymerase III (pol III) in some viruses [10]. The cleavage of pri-miRNAs releases a RNA hairpin intermediate (~70 nt) containing a characteristic 2 nt 3' overhang, named a premature miRNA (pre-miRNA), which is further processed to generate the 21~23 nt mature miRNA from its arm of ~70 nt imperfect stem-loop structure [11, 12].

Since microRNAs have been discovered and their role in gene expression regulation was established, it has been hypothesized that viruses could encode microRNA-like small RNAs as well, and these virus-encoded microRNA-like small RNAs were proposed to play important regulatory roles in viral immune evasion and systemic pathogenesis [13–15]. The size of viral encoded microRNA-like RNAs has a significant advantage given the tight constraints on viral genome size, which is also small enough to escape from the triggered host immune pathway. It was found that viral encoded microRNA-like small RNAs could downregulate the expression of host immune defense gene, resulting in increased viral replication or evasion from host immune surveillance [16, 17]. Until now, more than 60 viral microRNA-like small RNAs have been identified [18–24], most of which came from Herpes virues [25]. Only a small part of such RNAs was detected within Retrovirus, Adenovirus, and polyomavirus families [26–28].

Bioinformatics-driven prediction was an effective method to identify viral encoded micro-RNA-like small RNAs [21, 22]. In this study, the microRNA prediction program, VMir, was applied to scan the viral genomes for the presence of stem-loop structures in the pri- and premiRNAs and identify potential candidate stretches capable to form stable secondary stem-loop structures. Afterward, putative mature microRNA-like small RNAs were validated using MatureBayes [29]. The systemic prediction of the potential EBOV-encoded microRNA-like small RNAs along with their target genes on the genome-wide scale helps to further assess the function of microRNAs during viral infection and virus-host interactions in the EVD pathogenesis.

2. Methods

2.1. EBOV whole genome sequences and alignment

The full-length genome sequences of EBOV were retrieved from the genome browser at Ebola virus resource (http://www.ncbi.nlm.nih.gov/genome/viruses/variation/ebola/) and UCSC Ebola portal (https://genome.ucsc.edu/ebolaPortal/). MAFFT Multiple Sequence Alignment Software Version 7 were applied for the alignment of the EBOV genomes [30].

2.2. Bioinformatics prediction of the EBOV genome-encoded microRNA-like small RNAs

Briefly, the viral genome was scanned for stem-loop structures of miRNA precursor (premiRNA) using VMir [31] with default parameter settings (http://www.hpi-hamburg.de/ research/departments-and-research-groups/antiviral-defense-mechanism/software-down-

References

- Cenciarelli, O., Pietropaoli, S., Malizia, A., et al. Ebola virus disease 2013-2014 outbreak in west Africa: an analysis of the epidemic spread and response. *Int J Microbiol*. 2015, 769121 (2015).
- [2] Ksiazek, T.G. Filoviruses: Marburg and Ebola. Viral Infections of Humans. 14, 337–350 (2014).
- [3] World Health Organization. "Global Alert and Response (GAR): Situation Reports: Ebola Response Roadmap," 2015. Available at: http://www.who.int/csr/disease/ebola/ situation-reports/en/.
- [4] World Health Organization. "Ebola Situation Reports," 2014. Available at: http://apps.who.int/ebola/ebola-situation-reports.
- [5] Frieden, T.R., Damon, I., Bell, B.P., Kenyon, T. and Nichol, S. Ebola 2014 new challenges, new global response and responsibility. *N Eng J Med*. 371, 1117–1180 (2014).
- [6] Walker, N.F., Whitty, C.J. Tackling emerging infections: clinical and public health lessons from the West African Ebola virus disease outbreak, 2014–2015. *Clin Med.* 15, 457–460 (2015).
- [7] Alexander, K.A., Sanderson, C.E., Marathe, M., et al. What factors might have led to the emergence of Ebola in West Africa? *PLoS Negl Trop Dis*. 9(6), e0003652 (2015).
- [8] Turner, C. Ebola virus disease: An emerging threat. Nursing. 44, 68–69 (2014).
- [9] Kim, V. N., Han, J. and Siomi, M.C. Biogenesis of small RNAs in animals. *Nat Rev Mol Cell Biol.* 10, 128–139 (2009).
- [10] Grundhoff, A. and Sullivan, C.S. Virus-encoded microRNAs. Virology. 411, 325–343 (2011).
- [11] Brodersen P. and Voinnet, O. Revisiting the principles of microRNA target recognition and mode of action. *Nat Rev Mol Cell Biol.* 10, 141–148 (2009).
- [12] Carthew, R.W. and Sontheimer, E.J. Origins and mechanisms of miRNAs and siRNAs. *Cell.* 136, 642–655 (2009).
- [13] Seo, G.J., Chen, C.J. and Sullivan, C.S. Merkel cell polyomavirus encodes a microRNA with the ability to autoregulate viral gene expression. *Virology*. 383, 183–187 (2009).
- [14] Pfeffer, S., et al. Identification of virus-encoded microRNAs. Sicence. 304, 734–736 (2004).
- [15] Cullen, B.R. Viruses and microRNAs. Nat Genet. 38, S25–S30 (2006).
- [16] Kincaid, R.P. and Sullivan, R.P. Virus-encoded microRNAls: an overview and a look to the future. *Plos Pathog.* 8, e10033018 (2012). DOI: 10.1371/journal.ppat.1003018.

- [17] Lecellier, C.H., et al. A cellular microRNA mediates antiviral defense in human cells. *Science*. 308, 557–560 (2005).
- [18] Walz, N., Christalla, T., Tessmer, U. and Grundhoff, A. A global analysis of evolutionary conservation among known and predicted gammaherpesvirus microRNAs. J. Virol. 84, 716–728 (2010).
- [19] Sullivan, C.S., et al. SV40-encoded microRNAs regulate viral gene expression and reduce susceptibility to cytotoxic T cells. *Nature*. 435, 682–686 (2005).
- [20] Samols, M.A., Hu, J., Skalsky, R.L. and Renne, R. Cloning and identification of a microRNA cluster within the latency-associated region of Kaposi's sarcoma-associated herpesvirus. J. Virol. 79, 9301–9305 (2005).
- [21] Cui, C. Prediction and identification of herpes simplex virus 1-encoded microRNAs. J. Virol. 80, 5499–5508 (2006).
- [22] Sullivan, C.S., Grundhoff, A.T., Tevethia, S., Pipas, J.M. and Ganem, D. SV40-encoded microRNAs regulate viral gene expression and reduce susceptibility to cytotoxic T cell. *Nature*. 435, 682–686 (2005).
- [23] Cai, X., et al. Epstein-Barr virus microRNAs are evolutionarily conserved and differentially expressed. *PLoS Pathog.* 2, e23 (2006).
- [24] Liang, H. Identification of Ebola virus microRNAs and their putative pathological function. *Sci China Life Sci.* 57, 973–981 (2014).
- [25] Pfeffer, S., et al. Identification of microRNAs of the Herpesvirus family. *Nat. Methods*. 2, 269–276 (2005).
- [26] Omoto, S., et al. HIV-1 nef suppression by virally encoded microRNA. *Retrovirology*. 1, 44 (2004).
- [27] Bennasser, Y., et al. Evidence that HIV-1 encodes an siRNA and a suppressor of RNA silencing. *Immunity*. 22, 607–619 (2005).
- [28] Rosewick, N. Deep sequencing reveals abundant noncanonical retroviral microRNAs in B-cell leukemia/lymphoma. *Proc. Natl. Acad. Sci. USA*. 110, 2306–2311 (2013).
- [29] Gkirtzou, K., Tsamardinos, I., Tsakalides P. and Poirazi, P. MatureBayes: a probabilistic algorithm for identifying the mature miRNA within novel precursors. *Plos One.* 5, e11843 (2010). DOI: 10.1371/journal.pone.0011843.
- [30] Katoh, K. and Standley, D.M. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol.* 30, 772–780 (2013).
- [31] Li, S.C., Shiau, C.K. and Lin, W.C. Vir-Mir db: prediction of viral microRNA candidate hairpins. Nucleic Acids Res. 36, 184–189 (2008).
- [32] Jiang, P. MiPred: classification of real and pseudo microRNA precursors using random forest prediction model with combined features. *Nucleic Acids Res.* 35, 339–344 (2007).

- [33] Sethupathy, P., Corda, B. and Hatziqeorqiou, A.G. TarBase: a comprehensive database of experimentally supported animal microRNA targets. *RNA*. 12, 192–197 (2006).
- [34] Kanehisa, M. and Goto, S. KEGG: kyoto encyclopedia of genes and genomes. Nucleic Acids Res. 28, 27–30 (2000).
- [35] Huang, D.W., Sherman, B.T. and Lempicki, R.A. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat. Protoc.* 4, 44–57 (2009).
- [36] Mi, H., Muruganujan, A. and Thomas, P.D. PANTHER in 2013: modeling the evolution of gene function, and other gene attributes, in the context of phylogenetic trees. *Nucleic Acids Res.* 41, 377–386 (2013).
- [37] Mi, H., Muruganujan, A., Casaqrande, J.T. and Thomas, P.D. Large-scale gene function analysis with the PANTHER classification system. *Nat Protoc.* 8, 1551–1566 (2013).
- [38] Franceschini, A., et al. STRING v9.1: protein-protein interaction networks, with increased coverage and integration. *Nucleic Acids Res.* 41, 808–815 (2013).
- [39] Huang, D.W., Sherman, B.T. and Lempicki, R.A. Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. *Nucleic Acids Res.* 37, 1–13 (2009).
- [40] Bayarsaihan, D. Epigenetic mechanisms in inflammation. J. Dent. Res. 90, 9–17 (2011).
- [41] Hayden, M.S., West, A.P. and Ghosh, S. NF-kappaB and the immune response. Oncogene. 25, 6758–6780 (2006).
- [42] Bouwmeester, T. A physical and functional map of the human TNF-alpha/NF-kappa B signal transduction pathway. *Nat. Cell Biol.* 6, 97–105 (2004).
- [43] Xu, W. Ebola virus VP24 targets a unique NLS binding site on karyopherin alpha 5 to selectively compete with nuclear import of phosphorylated STAT1. *Cell Host Microbe*. 16, 187–200 (2014).
- [44] Kagoya, Y. Positive feedback between NF-kB and TNF-alpha promotes leukemiainitiating cell capacity. J. Clin. Invest. 124, 528–542 (2014).
- [45] Lee, R.E., Walker, S.R., Savery, K., Frank, D.A. and Gaudet, S. Fold change of nuclear NF-kB determines TNF-induced transcription in single cells. *Mol. Cell.* 53, 867–879 (2014).
- [46] Ramos, H.J. and Gale, M., Jr. RIG-I like receptors and their signaling crosstalk in the regulation of antiviral immunity. *Curr Opin Virol.* 1, 167–176 (2011).
- [47] Loo, Y.M. and Gale, M., Jr. Immune signaling by RIG-I-like receptors. *Immunity*. 34, 680–692 (2011).
- [48] Solis, M., et al. RIG-I-mediated antiviral signaling is inhibited in HIV-1 infection by a protease-mediated sequestration of RIG-I. *J Virol.* 85, 1224–1236 (2011).

- [49] Pichlmair, A., et al. RIG-I-mediated antiviral responses to single-stranded RNA bearing 5'-phosphates. *Science*. 314, 997–1001 (2006).
- [50] Gupta, A., et al. Anti-apoptotic function of a microRNA encoded by the HSV-1 latencyassociated transcript. *Nature*. 442, 82–85 (2006).
- [51] Satoh, T., et al. LGP2 is a positive regulator of RIG-I- and MDA5-mediated antiviral responses. *Proc Natl Acad Sci U S A*. 107, 1512–1517 (2010).
- [52] Shi, J. Identification and validation of a novel microRNA-like molecule derived from a cytoplasmic RNA virus antigenome by bioinformatics and experimental approaches. *Virol. J.* 11, 121:1–121:14 (2014).
- [53] Zuker, M. Mfold web server for nucleic acid folding and hybridization prediction. *Nucleic Acids Res.* 31, 3406–3415 (2003).
- [54] Parisien, M. and Major, F. The MC-fold and MC-sym pipeline infers RNA structure from sequence data. *Nature*. 452, 51–55 (2008).
- [55] Randall, R.E. and Goodbourn, S. Interferons and viruses: an interplay between induction, signalling, antiviral responses and virus countermeasures. J. Gen. Virol. 89, 1–47 (2008).
- [56] Kaul, D., Ahlawat, A. and Gupta, S.D. HIV-1 genome-encoded HIV1-MIR-H1 impairs cellular responses to infection. *Mol. Cell Biochem.* 323, 143–148 (2009).
- [57] Hussain, M. West Nile virus encodes a microRNA-like small RNA in the 3' untranslated region which up-regulates GATA4 mRNA and facilitates virus replication in mosquito cells. *Nucleic Acids Res.* 40, 2210–2223 (2012).
- [58] Hussain, M., Taft, R.J. and Asgari, S. An insect virus-encoded microRNA regulates viral replication. J. Virol. 82, 9164–9170 (2008).